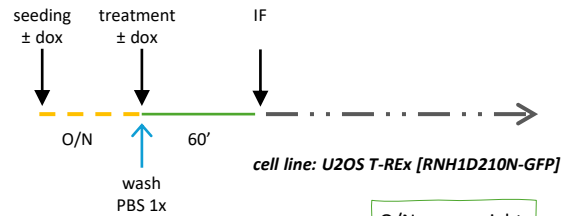


2. Monitoring the formation of R-loops in human cell

0 day

- seed cells 20 000 – 25 000 cells/cm²; O/N
- to be at approx. 70% confluence next day;
- treatment with dox



1st day

1 well ... 1 mL DMEM
... 2 mL 1xPBS

- ✿ pre-warm: DMEM, PBS
- ✿ cool down PBS (in fridge or on ice; approx. 4°)
- ✿ cool down 0.2 % Triton X-100 in PBS; (approx. 4°)
- ✿ cool down MeOH (-20°C)
- ✿ prepared all reagents and treatment

Keep cells on ice Cell culture room

- add 1 mL pre-warm DMEM to 6-well plate
- transfer CVs from plate to 6-well plate
- exchange DMEM for DMEM with treatment
- incubate (60 min; 37°C; 6% CO₂)
- wash cold PBS (2 mL) work in the lab on bench

wash
1. aspirate liquid
2. add washing solution
3. aspirate liquid
..... repeat
X. add desire liquid

treatment	c stock	c final	μL / 1 mL
dox	1 μg/mL	1 ng/mL	1 μL
APH	2 mM	2 μM	1 μL
CDDP	1 mM	10 μM	10 μL
CPT	100 μM	100 nM	1 μL

MM (Master mix)
in 15-mL falcon
dox 5 μL / 5 mL of DMEM
from that prepare individual treatment (in eppendorph)

Pre-extraction

- cold 0.2 % Triton X-100 in PBS (1mL; 10 min; on ice)
- wash with cold PBS (2 x 2 mL)

Fixation (work in the hood)

- 4 % (v/v) PFA in PBS (1mL; 15 min; r.t.)
- wash with PBS (3 x 2 mL)
- -20°C MeOH (1mL; 20 min; -20°C - freezer);
- wash with PBS (3 x 2 mL)

PFA
fixation via crosslinks

MeOH
dehydration and denaturation
for better access of PCNA epitope

Blocking

- 3% BSA in PBS (1mL; 20 min)

3% BSA ... 1.5 g ... 50 mL

- ✿ prepare wet chamber
- ✿ dilute primary antibody

Primary antibody

- dilute the antibodies in blocking solution
anti-PCNA (PC10) mouse 1:500
- 50 μL/CV
- place CVs on parafilm into wet chamber; cells facing up? ? ?
- incubation (1h; r.t.; wet chamber)

- wash with PBS (3 x 200 μL; 5min)

Secondary antibody

- dilute the antibodies in blocking solution in the one eppendorph
anti-mouse cy3 (555nm) 1:500
- DAPI 1:1000
- 50 μL/CV
- incubation (30 min; r.t.; wet chamber - dark)

???
secondary antibody have to be kept in dark from on

- wash with PBS (3 x 200 μL; 5min)
- dip to H₂O
- dip to tissue to remove excess of liquid
- air dry (approx. 30 min; r.t.; in dark ? ? ?)
- mount with Fluoromont 4 – 4.5 μl/slide (cells facing down? ? ?)
- air dry (r.t.; in dark ? ? ?)

2nd day

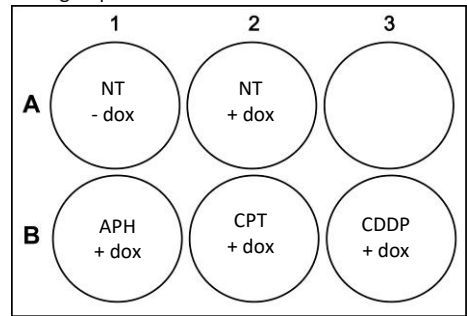
Microscopy

- DM6000 (objective lenses: 63x, 1.42 NA, oil)

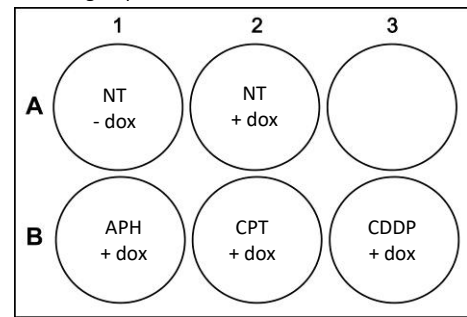
Note:

theory for fixation, blocking, primary and secondary ant is in protocol for Fibers assay (3)

First group



Second group



- ✿ **option:** incubation of primary antibodies O/N
- place CVs on parafilm into wet chamber; cells facing down? ? ?
- incubation (4°C; O/N; wet chamber)
- place CVs on parafilm into wet chamber; cells facing up? ? ?
- continue with wash etc. according to protocol

?? ? Choosing of secondary antibodies

⇒ based on their excitation and emission spectrum

- DAPI: emission 461 nm
- Green Fluorescent Protein (GFP): emission 488 nm fused with RNaseH1
- PCNA: has to have different emission (wavelengths), e.g. 555 nm, 647 nm

U2OS T-REx [RNH1D210N-GFP]

- derivative of the human osteosarcoma cell line U2OS
- expresses conditionally a catalytically-inactive form of RNase H1 (RNH1) fused C-terminally to green fluorescence protein (GFP)
- the RNH1D210N mutant can avidly bind to R-loops, but fails to degrade them and hence stabilizes these structures and can serve as a tool for their detection

Pre-extraction

- wash away soluble, non-chromatin-associated proteins before fixing the cell sample
- specifically bound or structural proteins by reducing background noise and improving the signal-to-noise ratio
- particularly for nuclear proteins

2. Monitoring the formation of R-loops in human cell

2nd-3rd day

ImageJ/Fiji

Protocol in detail in Analysis_Fiji_ImageJ

2. Monitoring the formation of R-loops in human cell

Treatment

CPT

- camptothecin
- fast, the most common
- inhibition of Top 1 → it leaves nicks; ↑ gH2AX; run off; SSB → DSB during replication
- ↑ negative torsion stress → ↑ R-loops serve as obstacle for replication
- (CPT works similar as depletion of Top1)

CDDP

- cisplatin
- adducts + oxidative stress; resolved by tranlesion synthesis and repriming

APH

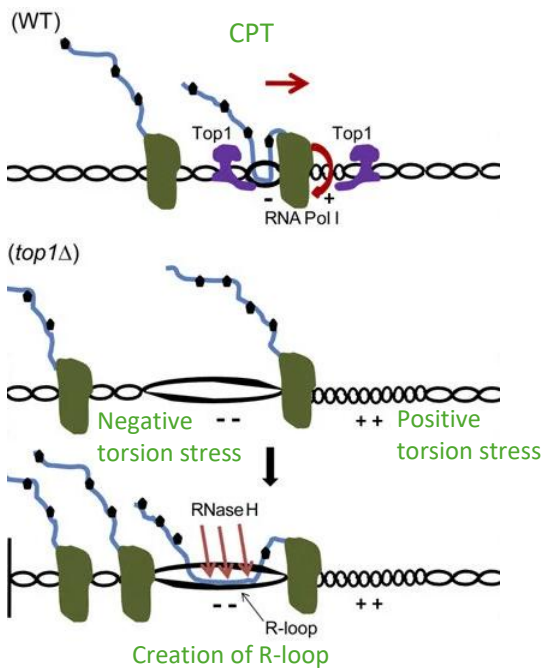
- aphidicolin
- inhibition of replicative polymerases;
- low concentration (0.2 μM) slow down; high block (2 μM)

dox

- doxycyclin
- induction of RNase H1 mutant – expression
- RNase H1 foci for R-loops visualization

PCNA

- proliferating cell nuclear antigen
- DNA clamp that acts as a processivity factor for DNA polymerase
- essential for replication
- can be used to distinguish of cells in S-phase

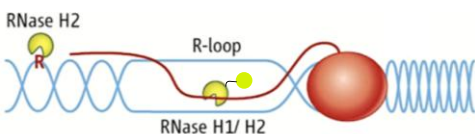


RNase-H1 wt - GFP:

- cell line: U2OS T-REx [RNH1WT-GFP]
- cuts R-loop
 - fluorescent for visualization of RNaseH1

RNase-H1 mutant - GFP:

- cell line: U2OS T-REx [RNH1D210N-GFP]
- doesn't cuts R-loop, only binds
 - fluorescent for visualization of R-loops



Tet ON system:
in the introduction

Reagent & solutions

PBS

- Phosphate-Buffered Saline
- 10x PBS ... stock; 10x concentrated
- 1x PBS ... as working solution, can be written as PBS in protocol

DMEM

Dulbecco's Modified Eagle Medium
a widely used basal medium for supporting the growth of many different mammalian cells

BSA

- bovine serum albumin

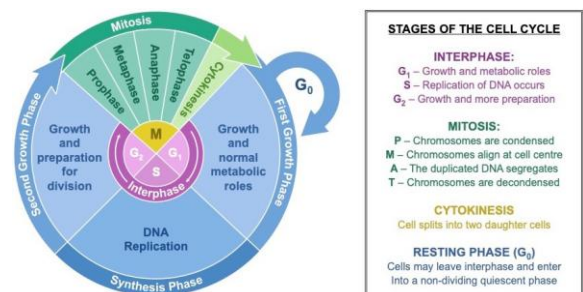
PFA or FA

- Paraformaldehyde

Triton-X

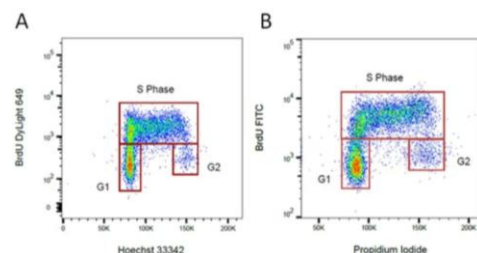
- a nonionic surfactant
- commonly used for membrane solubilization

Cell cycle

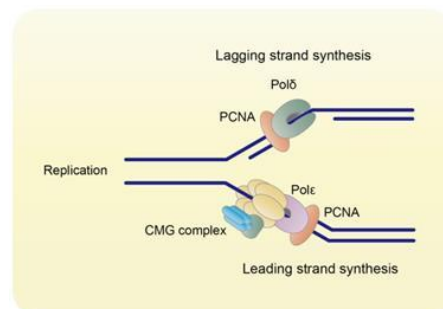


Visualization of cell cycle (S-phase)

- proteins or nucleotide analogues (e.g. thymidine: EdU, IdU, CldU, BrdU ...) which are specific for S-phase
- + DNA content (G1 ... N; G2 ... 2N) ... staining with DAPI or Hoechst or Propidium Iodide



PCNA



Source of images:

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DOI:10.1101/gad.573310

DOI:10.1083/jcb.201104003 <https://annualreport.nichd.nih.gov/2013/crouch.html>

<https://www.vce.bioninja.com.au/unit-one/area-of-study-1-cell-develo/cell-cycle.html>

<https://www.bio-rad-antibodies.com/flow-cytometry-proliferation-and-cell-cycle.html>

