

Basic processing in Fiji

Bio-formats import option (appears automatically for some image formats)

View stack with: Hyperstack → OK

Select and image → OK

ZOOM

Magnifying glass on or +/- → zoom in (mouse click) / zoom out (mouse click + Ctrl) / restore orig. size (mouse click + shift)

Open PROPERTIES

Image → Properties or Ctrl+Shift+P

Display METADATA

Image → Show Info or Ctrl+I

Open CHANNELS TOOL window (allows to simply change views – composite/color/greys)

Image → Color → Channels Tool or Ctrl+Shift+Z

Color more: LUT (upper bar) choose color: switch to the glow mode

Make Z-PROJECTION

Image → Stack → Z-project

Type
– max. intensity: nice contrast usually
– average

Change LUTs to "glow"

to see over/under exposure → good: do it close to the max. (blue)

Image → Lookup tables or LUT icon

To see all LUTs – Image → Color → Display LUTs

Open BRIGHTNESS/CONTRAST

Image → Adjust → Brightness/Contrast or Ctrl+Shift+C

use pref. min/max

⇒ ADJUST each channel of the image

(adjust min and max values, do not change gamma)

⇒ Press APPLY for each channel

Change LUTs back to desired color

Image → Lookup tables or LUT icon

→ Blue, green red/cyan, magenta → do not use green and red together

Switch to COMPOSITE using CHANNELS TOOL

Image → Color → Channels Tool or Ctrl+Shift+Z

SAVE the image as TIFF (stack of 3 grayscale images)

Add SCALE BAR

Analyze → Tools → Scale Bar

MAKE RGB (1 RGB image)

Image → Color → Stack to RGB

Image → Type → RGB color/RGB stack

Save the image as TIFF

File → Save as → Tiff

Processing all images in one file

OPEN

- Bio-format Import Option
- Hyperstack
- (open all series when compatible)
- Color mode: Colorized/(Composite → split channel)
- (Autoscale)

MACRO

→ record

→ create

Back to original size of image

(yellow square at left upper corner

Shift + magnifying glass (bookmark bar)

Saving all images – opened as concatenate

Adjust images as usual

a) Image → Type → RGB

File → Save as → Image as sequence

b) Image → Color → Split Channels

Image → Adjust → Brightness/Contrast

→ ! must be 'Apply', otherwise it is not apply does not work with export !

Plugins → Bio-format → Bio-format Exporter TIFF Write each time point to a separate file → Uncompressed or LWZ compression

c) Image → Adjust

Image → Color → Merge channels → File → Save as → Image as sequence

↔ for each channel do not: "Image Type RGB "

Other useful tools in Fiji

Look on [ORTHOGONAL VIEWS](#)

Image → Stack → Orthogonal views or CTRL+SHIFT+H

SPLIT CHANNELS

Image → Color → Split Channels

MERGE CHANNELS

Image → Color → Split Channels

Recommendation: check “keep source”

PLOT PROFILE

Draw line → Analyze → Plot profile or CTRL+K

[MAKE SUBSTACK](#) (define a list of slices or a range of slices with an increment)

Image Stack Tools Make Substack ...

[MAKE DIMENSIONALITY](#), try various options

Image → Hyperstack → Reduce Dimensionality

- isolate one time-frame, keep slices, keep channels
- isolate one slice, keep frames, keep channels
- isolate one channel ...

[CROP the image](#) (the original image down)

Draw a square → Image Crop or CTRL+Shift+X

[DUPLICATE a cut-out of the image](#) (the original image stays open)

[Adjust IMAGE SIZE](#) (re-define image height, width and/or depth)

Image Adjust Size

[Save as MOVIE](#)

(File → Save as → AVI ...)

[SYNCHRONIZE windows](#)

(Analyze → Tools → Synchronize windows → Synchronize All)

Final handling in Photoshop/Illustrator

Only for pictures for publication, not analysis

Open the TIFF image

[CROP a region and PASTE it to a new window](#)

Rectangular marquee tool → CTRL+C Ctrl+N Ctrl+V

Draw SCALE BAR

Rectangular tool set fixed pixels

or line with different stroke

→ must be done before resolution on 300 dpi (resampling)

merge layers afterwards

[Set IMAGE SIZE and RESOLUTION on 300 dpi](#) (check Resample image)

Image Image size or CTRL+ALT+I

Resample image (tick) than write resolution + contrast proportion

[Open a NEW FILE in Illustrator](#) (set desired size and change the color mode to RGB)

File → New

[COPY composite and each channel individually to Illustrator](#)

Ctrl+C and Ctrl+V

[Align images](#)

Window → Align

Scale bar issues

Resampling is better in

Scale bar:

e.g. $10 = 10\,000 \rightarrow 68.5 \text{ pix}$

ImageJ → Image → Show info down
Width: in microns (pixels)
Height: in microns (pixels)

Scale bar

LAS X: physical length = local pixels * voxel size = pixel size

Size of image in Illustrator: physical length * required scale bar = "real size of scale bar in illustrator"

e.g. $1398 * (0) * 203\,232 = 147.32 : 203\,232 * 15 = 10.87$

!!! Fiji - ImageJ: !!!

Analyze Tools Scale Bars Save as jpeg or png; if TIFF no scale bar

Gels and membrane analysis - quantification

Open images in ImageJ

Optional:

Adjust Brightness/contrast

We aware of control – everything has to be the same !!!!

Processing

- draw rectangle around first band
should be big enough to fit band and also for the rest
- Analyses → Gels → select first line (ctrl+1)
- Drag rectangle from the first line (with arrow, rectangle will duplicate) and label second, etc. the last line (! all rectangles have to have the same size !)
- Analyses → Gels → Plot lanes
- Graph: integration of peaks: use line tool (+Shift) or draw line with pencil
 - For WB (white background and black bands): area under {maybe above} just check peak
 - For ELFO gel (black background and white bands): area above peak, or invert colors and process as Wb membranes
- Magic wand tool → to select area under graph → Measure To see numbers copy to excel
- Results can be saved as Excel file

Optional

- Draw one rectangle for all bands integrate in one graph !!! Do not mix up the samples!!!
- Note: in some Fiji versions that doesn't work properly

Note

- There is many videos online on YouTube